

UNITED STATES PATENT APPLICATION

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FOR

CLONING, SEQUENCING AND EXPRESSION OF A GENE ENCODING AN
EUKARYOTIC AMINO ACID RACEMASE, AND DIAGNOSTIC, THERAPEUTIC,
AND VACCINATION APPLICATIONS OF PARASITE AND VIRAL MITOGENS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application is based on and claims the benefit of each of the following applications: U.S. Provisional Application S.N. 60/168,631, filed December 3, 1999 (attorney docket no. 03495.6044); U.S. Provisional Application S.N. 60/220,207, filed July 24, 2000 (attorney docket no. 3495.6054); and U.S. Provisional Application S.N. 60/221,117, filed July 27, 2000 (attorney docket no. 3495.6055). The entire disclosure of each of these applications is relied upon and incorporated by reference herein.

BACKGROUND OF THE INVENTION

This invention relates to the discovery of a new gene, which is the first isolated and cloned, that encodes an amino acid eukaryotic racemase. The invention covers, particularly, the Tc45 gene encoding a *Trypanosoma cruzi*-derived B-cell mitogen. The encoded protein also is a eukaryotic proline racemase. The invention also relates to a process of production of D-amino acid using an eukaryotic amino acid racemase. This invention also relates to the use of the protein encoded by the Tc45 gene to induce a protective immune response against *T. cruzi* infection in a human. This invention also relates to methods of using other parasite mitogens and viral mitogens for inducing protective immunity against the corresponding parasitic or viral infections in humans.

The process of production of an D-amino acid by using a L-amino acid source comprises the use of an eukaryotic amino acid racemase specific for the amino acid of interest, the said racemase being produced from a recombinant expression system containing a vector having a polynucleotide sequence encoding the said enzyme. In prokaryotic hosts, the racemases are known to be implicated in the

synthesis of D-amino acids and/or in the metabolism of L - amino acids. Therefore, the presence of free D - amino acids in tumors and in progressive autoimmune and degenerative diseases suggests the biological importance of eukaryotic amino acid racemases. It is well known that proteins or peptides containing D - amino acids are resistant to proteolysis by host enzymes. In addition, such proteins containing D - amino acids, at least one D - amino acid residue, can display antibiotic or immunogenic properties.

Isolation and characterization of molecules playing a key role in parasite metabolism, or in their interactions with the host immune defense, are fundamental for the development of rational strategies for vaccination and therapy. Attempts to provide effective immunity to parasites are limited by poor specific immune responses to parasite antigenic molecules in early phases of infection. Lymphocyte polyclonal activation is a generalized mechanism of immune evasion amongst pathogens¹. Such "parasite evasion" owes, at least in part, to the release of mitogenic or superantigenic moieties that inhibit host specific responses by triggering polyclonal, parasite non-specific lymphocyte activation. The resulting non-specific immune responses are associated with immunosuppression and autoimmunity, as observed in human and experimental infections by the protozoan parasite *Trypanosoma cruzi*, the etiological agent of Chagas disease²⁻⁶.

To date, there is no effective treatment or vaccine against *Trypanosoma cruzi* infection and Chagas disease pathology. Attempts to isolate immunodominant protective epitopes have failed¹. Using a mouse model of *T. cruzi* infection it has previously been shown that reduced levels of polyclonal lymphocyte responses correlate with resistance to infection and cardiopathy^{2, 7-9}. As we have suggested,

and has been demonstrated by Arala-Chaves³¹ for *Candida albicans* infections, mitogenic moieties can be used as vaccination targets to induce specific neutralization of the mitogen, thus aborting the microorganism "strategy" to deviate immune responses into non-specific polyclonal activation and immunosuppression. Understanding the mechanisms underlying "non-specific" lymphocyte activation may open the way for their neutralization, and thus allow for effective immune responses against infectious agents. There is a need in the art for a molecule that could be an appropriate target for such attempts.

Furthermore, there is a growing interest in the biological role of D-amino acids, either as free molecules or within polypeptide chains in human brain, tumors, anti-microbial and neuropeptides, as well as in "protein fatigue"³², suggesting widespread biological implications. Research on D-amino acids in living organisms has been hampered by their difficult detection. However, recent purification of a serine racemase from mammalian brain³³ indicates conservation throughout evolution. There also exists a need in the art for racemases that are specific for known compounds.

SUMMARY OF THE INVENTION

This invention aids in fulfilling these needs in the art. More particularly, this invention relates to the characterization of a parasite molecule implicated in polyclonal responses that may serve as a novel target for vaccination and therapy. After identifying a protein with B-cell mitogenic properties in culture supernatants of infective parasite forms, the corresponding gene was cloned and its genomic organization was characterized. The protein has been characterized as a cofactor independent proline racemase, with strong homology to the proline racemase

isolated from *Clostridium sticklandii*, thus providing the first report on an eukaryotic amino acid racemase gene.

In particular, this invention provides a purified peptide comprising an amino acid sequence (SEQ ID NOS: 1, 2, 3 and 4) encoded by the Tc45 gene. This invention also provides polypeptide fragments derived from SEQ ID NOS: 7, 8, 9, 10 and 11 containing at least 10 amino acids.

This invention additionally provides purified polynucleotides comprising the nucleic acid sequences of the Tc45 gene (SEQ ID NOS: 7, 8, 9, 10 and 11). This invention also provides nucleic acid fragments derived from SEQ ID NOS: 7, 8, 9, 10 and 11 containing 15 to 40 nucleotides.

Additionally, the invention includes a purified polynucleotide that hybridizes specifically under conditions of moderate stringency with a polynucleotide of SEQ ID NOS: 7, 8, 9, and 10.

SEQ. ID 7 represents the full nucleotide sequence encoding the *Trypanosoma cruzi* proline racemase and N-terminal signal sequence and the 5' and 3' flanking non-coding regions.

The SEQ ID 8 represents the full nucleotide sequence and its corresponding polypeptide sequences [including the N terminal signal sequence and the 3' non-coding flanking region] coding for a proline racemase of *T. cruzi*.

The construct as disclosed in SEQ ID 8 deleted of the 3' non-coding flanking region and inserted in the PET28 vector (NOVAGEN), transformed in *E. coli* DH5 α was deposited at the CNCM under the accession number I-2344.

A derived construct of I-2344 deleted of nucleotide sequence corresponding to the signal peptide coding sequences, which is described in SEQ ID 3, is used for the production of a recombinant active proline racemase in *E. coli*. The *E. coli* DH 5 α containing the plasmid with an insert of 239 base pairs deposited at CNCM under accession number I-2221, was obtained after amplification of the region by PCR... technique with the primers SEQ ID Nos. 12 and 13. The insert was cloned into p TOPO II commercialized by INVITROGEN and then transformed in *E. coli*.

The invention further includes polynucleotide fragments comprising at least 10 nucleotides capable of hybridization under conditions of moderate stringency conditions with any one of the nucleotide sequences enumerated above.

In another embodiment of the invention, a recombinant DNA sequence comprising at least one nucleotide sequence enumerated above and under the control of regulatory elements that regulate the expression of racemase activity in a host is provided.

The invention also includes a recombinant host cell comprising a polynucleotide sequence enumerated above or the recombinant vector defined above.

In still a further embodiment of the invention, a method of detecting parasitic strains that contain the polynucleotide sequences set forth above is provided.

Additionally, the invention includes kits for the detection of the presence of parasitic strains that contain the polynucleotide sequences set forth above.

The invention also contemplates antibodies recognizing peptide fragments or polypeptides encoded by the polynucleotide sequences enumerated above.

Still further, the invention provides for a screening method for active molecules for the treatment of infections due to parasites, particularly *T. Cruzi*, based on the detection of activity of these molecules on parasites.

This invention further provides an immunizing composition containing at least a purified protein, or a fragment thereof, capable of inducing an immune response *in vivo*. The immune response can be a mitogenic polyclonal immunoresponse *in vivo*. The immunizing composition is suitable for use against a parasite infection under sub-mitogenic doses.

This invention also provides a process to access the mitogenicity of a molecule called mitogen and the procedures to determine the sub-mitogenic dose suitable as an immunizing composition for use against a parasite infection.

A vaccine composition of the invention for use against a *T. cruzi* infection comprises a purified 38 to P45 kda protein or a fragment thereof .

A method of inhibiting an eukaryotic protein with an amino acid racemase activity according to the invention comprises treating a patient by administering an effective amount of a molecule that inhibits the eukaryotic protein. The parasite can be *T. cruzi*.

This invention also provides a process for screening a molecule capable of inhibiting the amino acid racemase activity of an eukaryotic protein comprising the steps of:

- contacting the purified eukaryotic racemase protein with standard doses of a molecule to be tested;
- measuring inhibition of racemase activity; and
- selecting the molecule.

BRIEF DESCRIPTION OF THE DRAWINGS

This invention will be more fully described with reference to the drawings in which:

Figure 1 depicts the proliferative activity of total spleen cells stimulated *in vitro* by a 45 kDa B-cell polyclonal activator isolated from parasite culture supernatants:

a. Proliferative activity of total spleen cells stimulated *in vitro* by increasing concentrations (below graph) of total proteins from culture supernatants of metacyclic trypomastigotes differentiated *in vitro*. Inset, Proliferation of total (■) or T cell-depleted (□) splenocytes in the presence of proteins from total culture supernatant c.p.m., counts per minute.

b. Proliferative activity of total spleen cells stimulated *in vitro* by HPLC pooled fractions at 24 h (■), 48 h (□) and 72 h (△). ³H-thymidine uptake (c.p.m., counts per minute) after 48 h in the presence of concavalin A and lipopolysaccharide was 32,000 and 3,500 counts per minute, respectively. To rule out the possibility of mitogenic effect due to lipopolysaccharide contamination in the samples, proliferation assays were also done using freshly recovered splenocytes from the 'lipopolysaccharide non-responder' mouse strain C3H/He; the same levels of proliferation were obtained.

c. 8% SDS-PAGE analysis of fractions 21-24 (silver staining). Left margin, molecular size markers. Arrow: correspond to a 45 kDa band.

Figure 2 shows the homology or similarity between the Tc45 protein (Tc) [SEQ ID NO:1] and the bacterial proline racemases *Clostridium sticklandii* (Cs) [SEQ ID NO.:5] and *Pseudomonas aeruginosa* (Pa) [SEQ ID NO.:6]. The computer-predicted signal peptide is indicated by a double arrow; peptide sequences obtained

by microsequencing appear underlined; peptides used for designing degenerate primers appear in italic characters. Proline racemase active sites are boxed; dashes indicate gaps generated for best fit.

Figure 3 shows the genomic organization and transcription of the *Tc45* gene.

a. Southern blot analysis of 5 µg of *T. cruzi* genomic DNA digested with indicated restriction enzymes and hybridized with a ³²P-labeled probe, including the TcPA45 coding sequence, is shown. Molecular weights are indicated.

b. is a Northern blot analysis of 20 µg of total RNA from epimastigotes forms hybridized with a ³²P-labeled probe as above. Molecular weights [Kb] are indicated.

c. mRNA expression of TcPA4S in different life stages of the parasite, shown by electrophoresis of gene fragments obtained by specific reverse transcription from total parasite RNA followed by PCR amplification using the sequences of the mini-exon (spliced leader) and R-300-45 primers and subsequent amplification of a 170-bp internal fragment. M1, M2 and M3, molecular size markers (sizes, left and right margins). First-strand cDNA reactions were done in the presence (+) or absence (-) of reverse transcriptase, to exclude the possibility of further PCR amplification of fragments due to genomic DNA contamination, C1 and C2, internal negative (no template) controls; E, epimastigote, T, trypomastigote and M, metacyclic.

Figure 4 shows the results of characterization of rTcPA45 activities *in vitro*.

a. 8 % SDS-PAGE gel of rTcPA45 (Coomassie blue staining).

b. Proliferative activity of total splenocytes (5x10⁴ cells/well) in the presence of increasing concentrations of rTcPA45 (µg/ml).

c. Percent racemisation of L-proline, D-proline, L-hydroxy-proline, D-hydroxy-proline substrates. Reaction conditions: 0.2 M Na-acetate/25mM β -mercaptoethanol buffer, pH6, rTcPA45 (3 μ g/ml), 30 min incubation at 37°C in 500 μ l. The reaction was stopped by incubating for 10 min at 80°C.

d. Percent inhibition of racemization of 80 mM L-proline in the presence of several inhibitors.

e. Percent racemization of L-proline (80 mM) as a function of pH (buffers used were 0.2 M: Na-acetate, K-phosphate and Tris-HCl and contained 25mM β -mercaptoethanol).

Figure 5 shows the nucleotide sequence [SEQ ID NO: 17] and peptide sequence [SEQ ID NO: 1] of TcPA45. The polypyrimidine rich region, splice leader acceptor sites, signal peptide, and polyadenylation site are indicated. The peptide sequences obtained by microsequencing of the native TcPA45 protein are underlined.

Figure 6 is a demonstration of a cytosolic proline racemase in epimastigote forms of *T. cruzi*.

Western blot: membranes containing in:

Lane 1: total epimastigote extract from 5×10^5 epimastigote forms

Lane 2: Epimastigote culture supernatant

Lane 3: Soluble fraction of epimastigote extract (cytosolic) from 5×10^5 epimastigote forms.

Lane 4: Insoluble fraction of epimastigote extract from 5×10^5 epimastigote forms

Lane 5: as in lane 1, from 10X more parasite forms (5×10^6 epimastigotes)

Lane 6: as in lane 3, from 10X more parasite forms (5×10^6 epimastigotes)

Lane 7: as in lane 2

Lane 8: as in lane 4, from 10X more parasite forms (5×10^6 epimastigotes)

Membranes were incubated with mouse polyclonal antibodies raised against the rTcPA45 protein (primary antibody). Second step reaction was done with goat anti-mouse IgG-horseradish peroxidase (human absorbed). Reactivities were developed by ECL-chemiluminescence Amersham kit. Figure 6 represents 10 seconds exposure of the film.

Figure 7 depicts the results of an ELISPOT assay. Spots correspond to immunoglobulin-producing B-cell (of a particular isotype) directed to the coated antigen (here: goat anti-mouse immunoglobulins or rTcPA45). See also Example 10 and Table 1.

Figure 8 depicts the results of DNA vaccination using various DNA constructs. See also Example 11.

Fig. 9 Characterization of rTcPA45 mitogenic activity.

Proliferative activity of total splenocytes obtained from athymic or euthymic mice in the presence of rTcPA45 or lipopolysaccharide, at 24 h (\square), 48 h (\angle) or 72 h (\blacksquare).

Fig. 10 Differential expression of rTcPA45 protein in the parasite.

a. Cellular localization of the Tc45 protein in different life stages of the parasite, shown by indirect immunofluorescence using polyclonal mouse serum against rTcPA45 followed by staining with the Alexa 488™ goat antibody against mouse IgG (H+L), F(ab')₂ fragment conjugate (bottom row), compared with control staining using the Alexa 488™ F(ab')₂ fragment conjugate alone (top left) or after incubation of the parasites with serum from chronically infected mice (Chronic serum).

b and c. Detection of Tc45 protein in total extracts of epimastigote (E), metacyclic (M) and trypomastigote (T) forms of the parasite, compared with recombinant rTcPA45 (R) protein, by western blot analysis. Arrows, calculated molecular weights for isoforms of the TcPA45 (R) protein.

d. Presence of the 39-kDa isoform of the Tc45 protein in total (Et), soluble (Ese) and insoluble (Emb) sonic extracts of non-infective epimastigote forms of the parasite, compared with its absence in culture supernatants (Ecs). Molecular sizes (b-d), left margins.

Fig. 11 Correlation between mitogenic and racemase activities.

a. Proliferative activity of total mouse splenocytes in the presence of rTcPA45 protein that is enzymatically active (rTcPA45) or lacking racemase activity by being heated (80°-rTcPA45), by long term storage at 4°C (Ina-rTcPA45), or by pre-incubation of rTcPA45 with pyrrole-Z carboxylic acid (rTcPA45 + 1 mM PCA), Iodoacetamide (rTcPA45 + 1mM IAA) or iodoacetate (rTcPA45 + 1 mM IAC) inhibitors, compared with 1 mM pyrrole-2-carboxylic acid 1 mMPCA) alone.

b. Competitive inhibition of rTcPA45-induced proliferative activity of total mouse lymphocytes by increasing concentrations of L- or D-proline substrates.

Controls, cultures of splenocytes with 50 mM L- or D-proline alone. c.p.m., counts per minute.

Figure 12 depicts the results of Western blot of mouse specific immune responses directed to mitogenic rTcPA45 following immunization. See also Example 15.

Figure 13 depicts two dose-response curves for two different hypothetical mitogens, A and B, in an assay of proliferative activity described hereinafter.

Figure 14 depicts the results of immunization according to the invention.

A. DNA vaccination. 8 week old BALB/c mice (5 mice/group) were injected 3 times (i.m., 50 µg of plasmid/femoral quadriceps) at 3 weeks interval with the following constructs: pcDNA3 vector alone as control, or pcDNA3 containing the full encoding sequence of the TcPA45 gene with (Long) or without (Short) the fragment encoding the signal peptide. Mice were challenged 4 weeks after the last injection with 10^4 infective forms of the parasite/mouse, and the parasitemia was scored during 35 days.

OBS. It is worth noting that BALB/c mice were treated by almost 2 months (9 weeks) to follow the vaccination protocol and were challenged at 21 weeks of age. Its is well known that mice of more than 9 weeks of age are naturally more resistant to the experimental infection with *Trypanosoma cruzi* and no morality is observed.

The results using this vaccination protocol revealed that 3 injections of pcDNA3 containing either the Short or Long encoding sequenvces of the TcPA45 gene, are able to reduce by more than 85% the parasitemia levels.

B. rTcPA45 injection. 6 week old BALB/c mice were injected intraperitoneally (i.p.) with 10ng of rTcPA45 and then boosted i.p. one week later with 50µg of rTcPA45. Mice were challenged 1 week after the boost with 10^4 infective forms of the parasite/mouse, and the parasitemia was scored during 30 days. Control mice did not receive any treatment before infection.

DETAILED DESCRIPTION

Having noted that proteins released from trypomastigote forms of *Trypanosoma cruzi* behave as polyclonal B-cell activators¹⁰, it was hypothesized that infective metacyclic forms would display an increased production of mitogenic molecules, thereby promoting such a mechanism of immune evasion. To investigate the release of proteins with B-cell mitogenic activity by *T. cruzi*, culture supernatants of *in vitro* differentiated metacyclic trypomastigotes in a protein-free defined medium were produced¹¹. Lymphocyte mitogenic activity in total concentrated culture supernatants was confirmed by lymphocyte proliferation assays.

Briefly, freshly recovered splenocytes from 8 week old male Balb/c mice seeded at a concentration of 5×10^4 cells/well were incubated for 24, 48 and 72 h in 5% FCS RPMI-1640 medium. A 16 h ^3H -thymidine pulse at 1µCi/well was performed before harvesting. ^3H -Thymidine uptake was determined in a beta-plate liquid scintillation counter (LKB-Wallac). All points were done in triplicate and the corresponding standard deviation calculated. For comparison, ^3H -thymidine uptake after 48 h in the presence of ConA (10 µg/ml) and LPS (5 µg/ml) was 32000 c.p.m. and 3500 c.p.m., respectively. T-cell depletion was performed by incubating freshly recovered spleen cells for 30 min at 37°C in the presence of anti-Thy 1.2 monoclonal antibody (Cedarlane) and rabbit complement (Cedarlane). In order to rule out a

possible mitogenic effect due to LPS contamination in samples, proliferation assays were also performed on freshly recovered splenocytes from the LPS non-responder mouse strain C3H/HeJ. Same levels of proliferation were verified. Balb/c mice were purchased from Charles River, France. C3H/HeJ mice were maintained in our animal facilities.

Proliferation is sustained over a 72 h period of culture in a dose dependent manner as shown in Figure 1a. Increasing concentrations ($\mu\text{g protein /ml}$) of total culture supernatant of *in vitro* differentiated metacyclic trypomastigotes are shown in this Fig. The inset shows proliferation of total or T-cell depleted splenocytes in the presence of 0.6 $\mu\text{g/ml}$ of total culture supernatant. As for other B-cell mitogens, the same level of proliferation was observed when supernatants were tested on T-cell depleted splenocytes (Figure 1a).

To identify the molecules responsible for mitogenic activity, the parasite proteins present in the metacyclic culture supernatants were fractionated by HPLC anion-exchanger chromatography, and the resulting fractions were tested as above. Fractions 22-24 (eluted at $368 \text{ mM} < \text{NH}_4\text{-acetate} < 467 \text{ mM}$) showed the highest ^3H -thymidine uptake at 24 hours. The results are shown in Figure 1b. 0.5 $\mu\text{g/ml}$ of HPLC pooled fractions were resuspended in RPMI.

Previous observations using DEAE-chromatography-purified parasite culture supernatants had shown that a protein fraction of 40-45 kDa was able to induce B-cell activation and proliferation *in vivo*. The estimation of this range of molecular weight was done more or less 10% and compared with standard molecular weight kit markers commercialized by BioLabs (USA).

SDS-PAGE analysis of the HPLC fractions revealed the presence of a protein with an apparent molecular weight around 45 kDa only in fractions 21-24 as shown in Figure 1c. (8% SDS-PAGE gel of fractions 21 to 24 (silver staining)). The isoelectric point of the protein was estimated between 4.5 and 5.0 by isoelectric focusing. These data indicated that a protein with B-cell mitogenic activity was present in fractions 21-24 eluted from the anionic matrix, and the 45 kDa protein band was thus selected as the main candidate.

To obtain peptide sequences from the 45 kDa protein, which was identified as Tc45, and allow for subsequent PCR-assisted cloning of a gene fragment, the 45 kDa protein band was isolated by SDS-PAGE, and its internal digestion and microsequencing were undertaken. Six peptides were isolated, sequenced, and were shown to have the following sequences: (W)⁴³IIK⁴⁶ [SEQ ID NO:18] ; I⁹⁰VTGSLP(D)I(S)G¹⁰⁰ [SEQ ID NO:19] ; A¹⁸³TNVPVVLDTAGLVR¹⁹⁸ [SEQ ID NO:20] ; V²⁴¹DIAFGGNF²⁴⁹ [SEQ ID NO:21] ; N³¹⁶VVIFGNR³²³ [SEQ ID NO:22] , and M³³⁸ATLYAK³⁴⁴ [SEQ ID NO:23] . These sequences are underlined in Figure 2.

Peptides 4 [SEQ ID NO:21] and 5 [SEQ ID NO:22] were selected for degenerate primer design on the basis of the relatively low level of degeneracy in their corresponding coding sequences. The sequences of these peptides are identified by italic characters in Figure 2.

Reverse transcription was performed on total RNA from *T. cruzi* trypomastigote forms using reverse degenerate primers for both peptides. The resulting cDNA was then used as a template for PCR amplification. From all possible combinations, when using in the PCR reaction forward primer for peptide 4 (SEQ ID No.13), reverse primer for peptide 5, and template cDNA synthesized with reverse primer for peptide 5 (SEQ ID No. 12), only one PCR product of 239 bp was shown to contain both primers after cloning and sequencing. The sequence analysis revealed that the fragment contained an unique open reading frame (ORF) flanked by peptides 4 [SEQ ID NO:21] and 5 [SEQ ID NO:22] coded in frame.

To obtain the full sequence of the *Tc45* gene, the ³²P-labelled 239 bp PCR product was used as a probe to screen a *Trypanosoma cruzi* clone CL-Brener lambda Fix II genomic library. Four independent positive phages were isolated. Restriction analysis and Southern blot hybridization revealed two types of patterns, each represented by two phages, suggesting that the *Tc45* gene is present in at least two copies per haploid genome.

The complete sequence of the *Tc45* gene and flanking regions (GenBank accession no. AF195522), revealed an ORF of 423 codons containing all sequenced peptides (Figure 2) [SEQ ID NO:1]. Computer analysis predicts a 29 amino acid signal peptide [SEQ ID NO: 3] (double arrowed in Figure 2) suggesting active secretion by *T. cruzi*. This is in agreement with the fact that the protein was purified from culture supernatants. A poly-pyrimidine rich region and probable trans-splice acceptor site is observed 56 and 7 base pairs (bp) upstream of the ATG codon, respectively. Interestingly, an alternative trans-splicing signal is present about 170 bp upstream of the second ATG codon within the coding region, which if used would

allow the expression of a truncated, non-secreted protein lacking 69 amino acids, which is SEQ ID No. 4. Polyadenylation may take place at positions 1442 or 1443, which are preceded by repeats of the triplet UUA, a motif found at the 3' end of other *T. cruzi* genes^{12,13}.

To investigate the genomic organization and transcription of the *Tc45* gene, Southern and Northern blot analyses were done. Figure 4 depicts the results of characterization of rTcPA45 activities. Fig. 4, part a, is an 8% SDS-PAGE gel of rTcPA45 (Coomassie blue staining). Part b depicts the proliferative activity of total splenocytes (5×10^4 cells/well) in the presence of increasing concentrations of rTcPA45 ($\mu\text{g/ml}$). Part c shows percent racemization of L-proline, D-proline, L-hydroxy-proline, D-hydroxy-proline substrates. Reaction conditions were: 0.2 M Na-acetate/25mM β -mercaptoethanol buffer, pH6, rTcPA45 (3 $\mu\text{g/ml}$), 30 min incubation at 37°C in 500 μl . The reaction was stopped by incubating for 10 min at 80°C. Fig. 4, part d, shows percent inhibition of racemization of 80 mM L-proline in the presence of several inhibitors. Part e shows percent racemization of L-proline (80 mM) as a function of pH (buffers used were 0.2 M: Na-acetate, K-phosphate and Tris-HCl and contained 25mM β -mercaptoethanol). Reactions were carried out for 30 min at 37°C and stopped for 5 min at 80°C. All reagents and inhibitors were purchased from Sigma. The shift in optical rotation was measured in a Polarimeter 241 MC (Perkin Elmer).

To investigate the genomic organization and transcription of the *Tc45* gene, we used Southern blot analysis; this indicated the presence of two gene copies per haploid genome. There are probably two homologous *Tc45* genes (Fig. 3a). Digestion with *Bam*HI and *Bgl*II producted two hybridizing bands, consistent with the

presence of two gene copies, as the probe has neither enzyme restriction site. High-molecular-weight DNA hybridized after probable partial digestion with *Sa*I, consistent with the absence of this site within the coding sequence covered by the probe. Both *Pst*I and *Taq*I cleaved within the probe and produced more than one hybridizing band per gene copy. Preliminary results indicated that they are located on different chromosomes (data not shown). Northern blot analysis on total RNA from epimastigotes showed a transcript of around 1.5 kb, as expected from the genomic sequence (Fig. 3b). We confirmed the presence of the Tc45 mRNA in different parasite forms by reverse transcription PCR using primers specific for the *Tc45* gene (Fig. 3c). Several point mutations were already identified in the sequence of the putative *Tc45-B* gene copy representative of the second phage type (Fig. 2), and further transcriptional and functional analyses of the alleles are underway.

To identify homologies with other genes, we compared the *Tc45-A* gene copy and protein sequence to several databases. There was homology (nucleotide sequence, 57.7%; IDENTITY amino-acid sequence, 52.4% IDENTITY) with the only proline racemase described¹⁴, an intracellular homodimeric protein isolated from *Clostridium sticklandii*. This enzyme catalyzes the interconversion between the L- and D-proline enantiomers, and its reaction mechanism has been studied extensively¹⁴. There was also homology with the translation of an ORF sequence from *Pseudomonas aeruginosa* in contig 53 of the unfinished *Pseudomonas* genome project (amino-acid sequence, 37.9% IDENTITY; *C. sticklandii* proline racemase and the translation of the *P. aeruginosa* ORF, 47% IDENTITY). The *C. sticklandii* proline racemase active site has been identified¹⁵ and is conserved in the Tc45 protein. Homology with both bacterial proteins started at amino acid 70 of Tc45 (Fig. 2). The

presence of the KIIK peptide (Fig. 2, underlining) in the Tc45 protein purified from parasite culture supernatants confirmed the presence of the extra N-terminal portion of the protein released by *T. cruzi* metacyclic forms.

More particularly, the SDS-PAGE analysis of the over-expressed and purified protein is shown in Fig. 4a. Using *in vitro* proliferation assays of naive murine spleen cells, recombinant protein rTc45 was shown to display a similar mitogenic activity to the one observed with the native protein fraction, purified from culture supernatants. Thus, rTcPA45 (for *T. cruzi* polyclonal activator 45) induces spleen lymphocyte proliferation, which increases with time over a 72 h period of culture (Figure 4b). Proliferation is dose dependent, with a bell-shaped response curve (starting from 0.8 µg/ml, and peaking at 50 µg/ml) typical of all mitogens described to date. rTcPA45 is indeed a T cell-independent polyclonal activator of B lymphocytes, as shown by the magnitude and increase of the proliferative response of total spleen lymphocytes obtained from athymic mice compared with the response of lymphocytes from euthymic individuals (Fig. 9). Injection of 50 µg rTcPA45 *in vivo* induced a 2 fold increase in spleen cell numbers by day 4, accompanied by an increase in numbers of immunoglobulin (Ig)-secreting B cells of the IgM, IgG2a, IgG2b and IgG3 isotypes (2.5 fold to 100x), while showing a complete lack of rTcPA45-specific Ig-secreting B cells, indicating the polyclonal B-cell mitogenicity of the protein (data not shown).

Injection of 50µg of rTcPA45 *in vivo* induces a 2-fold increase in spleen cell numbers by day 7 accompanied by 2.5 to 4 fold increase in numbers of Ig-secreting B cells of IgM, IgG2a, and IgG2b isotypes, while showing a complete lack of Ig-secreting B cells directed to TcPA45 demonstrating the polyclonal B-cell mitogenicity of the protein as shown in Example 10, Table I.

To confirm that the TcPA45 protein is indeed a proline racemase, *in vitro* biochemical assays were performed to measure the shift in optical rotation of either L- or D-proline substrates. As can be seen in Figure 4c, rTcPA45 racemises both L- and D-proline, but not L- or D- hydroxy-proline, nor any other natural L-amino acids. Such rTcPA45 racemase activity is co-factor independent, notably of pyridoxal phosphate, and thus closely resembles the *C. sticklandii* proline racemase¹⁴.

Furthermore, the rTcPA45 enzymatic activity is inhibited to different extents by the presence of previously described inhibitors¹⁴, such as maleic acid, iodoacetamide, iodoacetate, and pyrrole-2-carboxylic acid (Figure 4d). Interestingly, rTcPA45 proline racemase activity is maximal at pH 6 (Figure 4e), two units lower than that of the bacterial enzyme¹⁴. The optimal temperature for enzymatic activity is 37°C, and the enzyme is inactivated by 10 min heating at 80°C (data not shown).

To analyze the cellular localization of the parasite TcPA45, we used immunofluorescence experiments with a polyclonal serum raised against rTcPA45. Whereas serum from chronically infected mice stained trypomastigote cells uniformly, rTcPA45 specific antibodies stained mostly the cytoplasm of epimastigote forms but not the nucleus or the kinetoplast (Fig. 10a). *In vitro*-differentiated metacyclic forms showed a less intense and more diffuse pattern of cytoplasmic staining than did epimastigotes. However, bloodstream trypomastigote forms were strongly labeled at the flagellar pocket and the anterior and posterior ends of the parasite, and lightly along the flagellum and cytoplasm. These experiments substantiate the hypothesis that *T. cruzi* has an intracellular form of the proline racemase and the secretion might only take place in the infective forms. Western blot analysis of cell extracts of the parasite confirmed that the TcPA45 protein was present in different developmental

stages (Fig. 10b). We detected a TcPA45 protein around 39 kDa in molecular mass in epimastigotes (non-infective insect forms) and 41.5 kDa in infective metacyclic trypomastigotes, compared with the computer-predicted molecular masses of 43.4 kDa and 38 kDa, respectively, for a secreted and a non-secreted form of the protein (Fig. 10c). Western blot analysis of the non-infective epimastigote cell stage showed Tc45 proline racemase mostly in the soluble cellular fraction, only weakly in the cellular insoluble fraction and absent from culture medium (Fig. 10d).

To confirm the relationship, if any, between the enzymatic and the mitogenic activities of rTcPA45, we used *in vitro* proliferation assays with active rTcPA45 and different forms of the inactivated enzyme. Unexpectedly, mitogenic activity was abolished when rTcPA45 was inactivated by being heated or by long storage at 4 °C, or whenever enzymatic inhibition was achieved by pre-incubation of the protein with specific (pyrrole-2-carboxylic acid) or nonspecific (iodoacetamide and iodoacetate) inhibitors of proline racemase (Fig. 11a). Mitogenic activity was also affected considerably by supplementation of the cultures with increasing amounts of L- or D-proline, in a dose-dependent manner, indicating that competitive inhibition occurred in the presence of specific substrates (Fig. 11b). There was no cell proliferation when lymphocytes were cultured in the presence of 50mM L- or D-proline alone (Fig. 11b). Furthermore, mitogenic activity due to another B-cell mitogen was unaffected by the inhibitors or substances (data not shown). Although the antibodies against TcPA45 raised against the recombinant protein were not able to inhibit racemization or to neutralize mitogenic activity *in vitro*, and thus cannot be used to support a link between these activities, the results indicate that a free and intact active site of the rTcPA45 protein is necessary to allow mitogenicity.

This is the first description of an amino acid racemase gene in an eukaryotic organism. Thus, this invention relates to the biochemical isolation, cloning, and molecular characterization of a B-cell mitogen released by *Trypanosoma cruzi*. Unexpectedly, this is also the first description of a racemase in a parasite.

In bacteria, amino acid racemases are cytoplasmic proteins participating in metabolic processes or in the synthesis of post-translationally modified peptides¹⁵. It is known that *T. cruzi* can use L-proline as a major carbon source¹⁷, possibly through a D-proline intermediate¹⁸. In this case, however, one might expect to find a cytosolic proline racemase. Indeed, we have identified a cytosolic proline racemase in epimastigote forms of *T. cruzi* as shown in Figures 6 and 10(b). This racemase presents a molecular weight of 38 kd in 10% SDS Page by using a standard molecular weight kit commercialized by BioLABS (USA). The estimation of the molecular weight is done more or less 10% around 38 kda. It is well established that proteins bearing D- amino acids are highly resistant to eukaryotic proteases. Thus, it might also be possible that the parasite uses the racemization mechanism during metacyclogenesis to synthesize and express, on its surface, proteins containing D-proline, therefore ensuring a certain degree of resistance to host-induced proteolytic mechanisms during cell invasion. Interestingly, *T. cruzi* differentiation from epimastigote to trypomastigote is induced in the presence of L-proline at pH 6 in the insect's gut¹⁹ as well as during *in vitro* metacyclogenesis¹¹, and the proline racemase activity of the TcPA45 protein might be involved in this process.

Figure 2 shows homology between the Tc45 protein (Tc) and the bacterial proline racemases *Clostridium sticklandii* (Cs) and *Pseudomonas aeruginosa* (Pa). Identical residues appear in bold characters; the computer-predicted signal peptide is indicated by a double arrow; peptide sequences obtained by microsequencing appear underlined; peptides used for designing degenerate primers appear in italic characters. Proline racemase active sites are boxed; dashes indicate gaps generated for best fit.

The homology of TcPA45 protein sequence with the *Clostridium sticklandii* proline racemase starts at amino acid 70 (second methionine in the ORF of TcPA45 gene) allowing the speculation that *T. cruzi* has acquired an extra 70 amino acid sequence N-terminal to the ancestral proline racemase (comprising a signal peptide for secretion) gaining the ability to produce either a cytosolic or a secreted form of the protein by differential trans-splicing of the mRNA. In addition, this process might be differentially regulated at distinct developmental stages of the parasite. In this context, it could then be hypothesised that epimastigote forms produce a cytosolic protein, while infective trypomastigotes secrete the same protein that activates B-cell polyclonal responses.

Interestingly, the disruption of alanine racemase and the D-amino acid aminotransferase genes of *Listeria monocytogenes* results in the inability of the bacteria to grow within the eukaryotic host cells²⁴. Both these gene products are involved in the synthesis of D-alanine that is required for the production of a mucopeptide component of the cell walls of virtually all bacteria. It remains to be investigated whether or not such bacterial molecules are B-cell mitogens. Accordingly, working with random amino acid polymers, Sela and co-workers have

established that multichain polypeptides composed of D-amino acids induce antibody responses in a T-cell independent manner^{25, 26}, a property that can be interpreted as equivalent to B-cell mitogenicity^{27, 28}. Resistance of such D-amino acid peptides to degradation by host enzymes²⁹ could also explain the persistence of polyclonal responses (and immunosuppression)³⁰, even if parasite production of the mitogen would be transient at the start of infection.

Thus, novel polynucleotides corresponding to the Tc45 gene from CL strain (representative of lineage *T. cruzi* II) *T. Cruzi* have been isolated and sequenced. The presence of the Tc45 gene has also been demonstrated in a representative strain of another major lineage of *T. cruzi* [strain DM 28c, lineage *T. cruzi* I]. For a review on recommendations on *T. cruzi* strain nomenclature see: Mem. Institut. OSW. Cruz, Vol. 94, Suppl. 1, page 429-432, 1999. These polynucleotides include SEQ ID NOS: 7, 8, 9, 10, and 11. By "polynucleotides" according to the invention is meant the sequences referred to as SEQ ID NOS: 7, 8, 9, 10, and 11, and the complementary sequences and/or the sequences of polynucleotides that hybridize to the referred sequences under conditions of moderate stringency. The moderate stringency conditions are defined as washing conditions in 2 x SSC at 55°C, and hybridization operated in 5 X SSC at 55°C.

By "active molecule" according to the invention is meant a molecule capable of inhibiting the activity of the purified recombinant or native polypeptide as defined in the present invention.

Thus, the polynucleotides of SEQ ID NO: 7 and its fragments can be used as probes or to select nucleotide primers notably for an amplification reaction, such as the amplification reactions further described. PCR is described in the U.S. Patent

No. 4,683,202 granted to Cetus Corp. The amplified fragments may be identified by agarose or polyacrylamide gel electrophoresis, or by a capillary electrophoresis, or alternatively by a chromatography technique (gel filtration, hydrophobic chromatography, or ion exchange chromatography). The specificity of the amplification can be ensured by a molecular hybridization using as nucleic acid probes the polynucleotides derived from SEQ ID NO: 7 and its fragments, oligonucleotides that are complementary to these polynucleotides or fragments thereof, or their amplification products themselves.

Amplified nucleotide fragments are useful as probes in hybridization reactions in order to detect the presence of one polynucleotide according to the present invention or in order to detect the presence of a parasite of *T. Cruzi* strain carrying genes encoding racemase activity, in a biological sample. This invention also provides the amplified nucleic acid fragments ("amplicons") defined herein above. These probes and amplicons can be radioactively or non-radioactively labeled, using for example enzymes or fluorescent compounds.

Preferred nucleic acid fragments that can serve as primers according to the present invention are the following:

5'TTICCRAADAT|ACIACGTT3' [SEQ ID NO: 12]

5'ATHGCITTYGGIGGIAAYTTT3' [SEQ ID NO: 13]

5'TTICCRAADATIACIACGTT3' [SEQ ID NO: 14]

5'CTCTCCCATGGGGCAGGAAAAGCTTCTG3' [SEQ ID NO: 15]

5'CTGAGCTCGACCAGATCTATCTGC3' [SEQ ID NO: 16].

The primers can also be used as oligonucleotide probes to specifically detect a polynucleotide according to the invention.

Other techniques related to nucleic acid amplification can also be used alternatively to the PCR technique. The Strand Displacement Amplification (SDA) technique (Walker et al., 1992) is an isothermal amplification technique based on the ability of a restriction enzyme to cleave one of the strands at a recognition site (which is under a hemiphosphorothioate form), and on the property of a DNA polymerase to initiate the synthesis of a new strand from the 3' OH end generated by the restriction enzyme, and on the property of this DNA polymerase to displace the previously synthesized strand being localized downstream.

The SDA amplification technique is more easily performed than PCR (a single thermostated water bath device is necessary), and is faster than the other amplification methods. Thus, the present invention also comprises using the nucleic acid fragments according to the invention (primers) in a method of DNA or RNA amplification, such as the SDA technique. The polynucleotides of SEQ ID NO: 7 and its fragments, especially the primers according to the invention, are useful as technical means for performing different target nucleic acid amplification methods, such as:

- TAS (Transcription-based Amplification System), described by Kwoh et al. in 1989;
- SR (Self-Sustained Sequence Replication), described by Guatelli et al. in 1990;
- NASBA (Nucleic acid Sequence Based Amplification), described by Kievitis et al. in 1991; and
- TMA (Transcription Mediated Amplification).

The polynucleotides of SEQ ID NO: 7 and its fragments, especially the primers according to the invention, are also useful as technical means for performing methods for amplification or modification of a nucleic acid used as a probe, such as:

- LCR (Ligase Chain Reaction), described by Landegren et al. in 1988 and improved by Barany et al. in 1991, who employ a thermostable ligase;
- RCR (Repair Chain Reaction), described by Segev et al. in 1992;
- CPR (Cycling Probe Reaction), described by Duck et al. in 1990; and
- Q-beta replicase reaction, described by Miele et al. in 1983 and improved by Chu et al. in 1986, Lizardi et al. in 1988, and by Burg et al. and Stone et al. in 1996.

When the target polynucleotide to be detected is RNA, for example mRNA, a reverse transcriptase enzyme can be used before the amplification reaction in order to obtain a cDNA from the RNA contained in the biological sample. The generated cDNA can be subsequently used as the nucleic acid target for the primers or the probes used in an amplification process or a detection process according to the present invention.

Nucleic acid probes according to the present invention are specific to detect a polynucleotide of the invention. By "specific probes" according to the invention is meant any oligonucleotide that hybridizes with the polynucleotide of SEQ ID NO: 7, and which does not hybridize with unrelated sequences. Preferred oligonucleotide probes according to the invention are SEQ ID NOS:12, 13, 14, 15, and 16.

In a specific embodiment, the purified polynucleotides according to the present invention encompass polynucleotides having at least 80% identity in their nucleic acid sequences with polynucleotide of SEQ ID NO: 7 or fragments thereof. By percentage of nucleotide identity according to the present invention is intended a

percentage of identity between the corresponding bases of two homologous polynucleotides, this percentage of identity being purely statistical and the differences between two homologous polynucleotides being located at random and on the whole length of said polynucleotides. The calculation was made according to the software GCG and the program "gap."

The oligonucleotide probes according to the present invention hybridize specifically with a DNA or RNA molecule comprising all or part of the polynucleotide of SEQ ID NO: 7 under stringent conditions. As an illustrative embodiment, the stringent hybridization conditions used in order to specifically detect a polynucleotide according to the present invention are advantageously the following:

Prehybridization and hybridization are performed as follows in order to increase the probability for heterologous hybridization:

The prehybridization and hybridization are done at 50°C
in a solution containing 5 X SSC and 1 X Denhardt's
solution.

The washings are performed as follows:

2 X SSC at 60°C 3 times during 20 minutes each.

The non-labeled polynucleotides or oligonucleotides of the invention can be directly used as probes. Nevertheless, the polynucleotides or oligonucleotides are generally labeled with a radioactive element (^{32}P , ^{35}S , ^3H , ^{125}I) or by a non-isotopic molecule (for example, biotin, acetylaminofluorene, digoxigenin, 5-bromodesoxyuridin, fluorescein) in order to generate probes that are useful for numerous applications. Examples of non-radioactive labeling of nucleic acid

fragments are described in the French Patent No. FR 78 10975 or by Urdea et al. or Sanchez-Pescador et al. 1988.

Other labeling techniques can also be used, such as those described in the French patents 2 422 956 and 2 518 755. The hybridization step may be performed in different ways (Matthews et al. 1988). A general method comprises immobilizing the nucleic acid that has been extracted from the biological sample on a substrate (nitrocellulose, nylon, polystyrene) and then incubating, in defined conditions, the target nucleic acid with the probe. Subsequent to the hybridization step, the excess amount of the specific probe is discarded, and the hybrid molecules formed are detected by an appropriate method (radioactivity, fluorescence, or enzyme activity measurement).

Advantageously, the probes according to the present invention can have structural characteristics such that they allow signal amplification, such structural characteristics being, for example, branched DNA probes as those described by Urdea et al. in 1991 or in the European Patent No. 0 225 807 (Chiron).

In another advantageous embodiment of the present invention, the probes described herein can be used as "capture probes", and are for this purpose immobilized on a substrate in order to capture the target nucleic acid contained in a biological sample. The captured target nucleic acid is subsequently detected with a second probe, which recognizes a sequence of the target nucleic acid that is different from the sequence recognized by the capture probe.

The oligonucleotide fragments useful as probes or primers according to the present invention can be prepared by cleavage of the polynucleotide of SEQ ID NO: 7 by restriction enzymes, as described in Sambrook et al. in 1989. Another

appropriate preparation process of the nucleic acids of the invention containing at most 200 nucleotides (or 200 bp if these molecules are double-stranded) comprises the following steps:

- Synthesizing DNA using the automated methods, such as beta-cyanethylphosphoramidite described in 1986;
- cloning the thus obtained nucleic acids in an appropriate vector; and
- purifying the nucleic acid by hybridizing to an appropriate probe according to the present invention.

A chemical method for producing the nucleic acids according to the invention, which have a length of more than 200 nucleotides (or 200 bp if these molecules are double-stranded), comprises the following steps:

- Assembling the chemically synthesized oligonucleotides, which can have different restriction sites at each end;
- cloning the thus obtained nucleic acids in an appropriate vector; and
- purifying the nucleic acid by hybridizing to an appropriate probe according to the present invention.

The oligonucleotide probes according to the present invention can also be used in a detection device comprising a matrix library of probes immobilized on a substrate, the sequence of each probe of a given length being localized in a shift of one or several bases, one from the other, each probe of the matrix library thus being complementary to a distinct sequence of the target nucleic acid. Optionally, the substrate of the matrix can be a material able to act as an electron donor, the detection of the matrix positions in which hybridization has occurred being subsequently determined by an electronic device. Such matrix libraries of probes and methods of specific detection of a target nucleic acid are described in European patent application No. 0 713 016, or PCT Application No. WO 95 33846, or also PCT Application No. WO 95 11995 (Affymax Technologies), PCT Application No. WO 97 02357 (Affymetrix Inc.), and also in U.S. Patent No. 5,202,231 (Drmanac), said patents and patent applications being herein incorporated by reference.

The present invention also pertains to a family of recombinant plasmids containing at least a nucleic acid according to the invention. According to an advantageous embodiment, a recombinant plasmid comprises a polynucleotide of SEQ ID NO: 7 or nucleic acid fragment thereof. More specifically, the following plasmids are part of the invention:

DH5[alpha] - pTc45 MIT (1335 bp) and

Dh5[α] - pTc45 MIT (239 bp).

A suitable vector for the expression in bacteria, and in particular in *E. coli*, is pET-28 (Novagen), which allows the production of a recombinant protein containing a 6xHis affinity tag. The 6xHis tag is placed at the C-terminus or N-terminus of the recombinant polypeptide. The purified racemase is obtained by expression in *E. coli*

transformed with pET-28 containing the insert of the plasmid corresponding to the CNCM No. I-2344 deleted of the sequence of the signal peptide as shown on SEQ ID NO: 9 and including the six C terminal histidine residues. The expression of pET-28 with the insert was induced by IPTG (1 millimolar) overnight at 20°C resulting in a soluble recombinant racemase according to the invention. This racemase has a molecular weight of 45 kda in 8% SDS PAGE gel (see Figure 4a) compared with standard molecular weight kit markers (BioLABS). After lysis of the bacterial cells by a French Press, followed by centrifugation (2000 g during 15 minutes), the recombinant protein was purified from the supernatant using nickel IMAC chromatography. (commercialized by PHARMACIA) and eluted in 0.5 molar imidazol buffer. The yield is between 10 to 40 mg of protein for one liter of bacterial culture at 1.OD density. The OD at the beginning of the induction of the expression in the recombinant bacterial was comprised between 0.6 to 1 OD. In the culture conditions as disclosed above, the majority of the recombinant protein produced is in a soluble form and they are not favorable for the expression of the proteins of the bacterial host. The estimation of the molecular weight of the purified recombinant protein is done more or less 10% around 45 kda.

The polypeptides according to the invention can also be prepared by conventional methods of chemical synthesis, either in a homogenous solution or in solid phase. As an illustrative embodiment of such chemical polypeptide synthesis techniques, the homogenous solution technique described by Houbenweyl in 1974 may be cited.

The polypeptides of the invention are useful for the preparation of polyclonal or monoclonal antibodies that recognize the polypeptides (SEQ ID NOS: 1, 2, 3, and

4) or fragments thereof. The monoclonal antibodies can be prepared from hybridomas according to the technique described by Kohler and Milstein in 1975. The polyclonal antibodies can be prepared by immunization of a mammal, especially a mouse or a rabbit, with a polypeptide according to the invention, which is combined with an adjuvant, and then by purifying specific antibodies contained in the serum of the immunized animal on a affinity chromatography column on which has previously been immobilized the polypeptide that has been used as the antigen.

Consequently, the invention is also directed to a method for detecting specifically the presence of a polypeptide according to the invention in a biological sample. The method comprises:

- a) bringing into contact the biological sample with an antibody according to the invention; and
- b) detecting antigen-antibody complex formed.

Also part of the invention is a diagnostic kit for *in vitro* detecting the presence of a polypeptide according to the present invention in a biological sample. The kit comprises:

- a polyclonal or monoclonal antibody as described above, optionally labeled; and
- a reagent allowing the detection of the antigen-antibody complexes formed, wherein the reagent carries optionally a label, or being able to be recognized itself by a labeled reagent, more particularly in the case when the above-mentioned monoclonal or polyclonal antibody is not labeled by itself.

Indeed, the monoclonal or polyclonal antibodies according to the present invention are useful as detection means in order to identify or characterize a *T. Cruzi* strain carrying TC45 genes.

The invention also pertains to:

- A purified polypeptide or a peptide fragment having at least 10 amino acids, which is recognized by antibodies directed against a polynucleotide or peptide sequence according to the invention.

- A polynucleotide comprising the full length coding sequence of the Tc45 gene sequence according to the invention.

- A monoclonal or polyclonal antibody directed against a polypeptide or a peptide fragment encoded by the polynucleotide sequences according to the invention.

- A method of detecting the presence of parasite harboring the polynucleotide sequences according to the invention in a biological sample comprising:

- a) contacting DNA or RNA of the biological sample with a primer or a probe according to the invention, which hybridizes with a nucleotide sequence;
- b) amplifying the nucleotide sequence using said primer or said probe; and
- c) detecting the hybridized complex formed between said primer or probe with the DNA or RNA.

A kit for detecting the presence of a parasite harboring the polynucleotide sequences according to the invention in a biological sample, comprises:

- a) a polynucleotide probe according to the invention; and

- b) reagents necessary to perform a nucleic acid hybridization reaction.

A method of screening active molecules for the treatment of the infections due to a parasite, comprises the steps of:

- a) bringing into contact a parasite containing the polynucleotide sequences according to the invention with the molecule; and
- b) measuring an activity of the active molecule on the parasite.

An *in vitro* method of screening for an active molecule capable of inhibiting a polypeptide encoded by the polynucleotide sequences according to the invention, wherein the inhibiting activity of these molecules is tested on at least said polypeptide, comprises the steps of:

- a) providing a polypeptide according to the invention;
- b) contacting the active molecule with said polypeptide;
- c) testing the capacity of the active molecules, at various concentrations, to inhibit the activity of the polypeptide; and
- d) choosing the active molecule that provides an inhibitory effect of at least 80 % on the activity of the said polypeptide.

A test for screening the inhibiting activity of a molecule, for example, a new substrate analogue or a new antiparasitic agent, can comprise the following steps:

A suitable test for testing an active molecule inhibiting the polypeptide according to the invention is performed as follows:

The recombinant amino acid purified or native racemase is diluted in sodium acetate buffer or Tris or phosphate or Hepes buffer at 3 micrograms per 500 microliters in the

presence of 20 millimolar of beta mercapto ethanol and 10 to 80 millimolar of L or D substrate and containing various concentrations of active molecule to be tested. This reaction is incubated for 30 minutes at 37°C and stopped by heating at 80°C. Variations in optical rotation are measured by a polarimeter.

Another embodiment of this invention provides a method for inhibiting the activity of a parasite *in vivo*. The method comprises administering to a host a parasite mitogen, which is capable of exhibiting a protective effect, a curative effect, or preventing transmission of a parasite in the host. The parasite mitogen is administered to the host in an amount sufficient to prevent or at least inhibit infection *in vivo* or to prevent or at least inhibit spread of the parasite *in vivo*. These effects are achieved by administering the parasite mitogen to the host in a sub-mitogenic amount, which is preferably sufficient to induce a protective response against the parasite in the host.

The parasite mitogen employed in this invention is distinguished from an "antigen", which is a substance that induces an immune response, such as a complete antigen that both induces an immune response and reacts with the product of the response, or an incomplete antigen (hapten) that cannot induce an immune response by itself, but can react with the products of an immune response when complexed to a complete antigen (carrier). The parasite mitogens of the present invention are thus unlike antigens, which require processing and presentation, such as (1) uptake of the antigen by antigen presenting cells (APCs); (2) internalization of the antigen in intracellular vesicles; (3) intracellular processing, which may include

the unfolding of a protein and/or partial proteolysis, with generation of immunogenic peptides; (4) binding of peptides to class II MHC molecules to form a bimolecular complex recognized by T cells; and (5) transport to, and display of, the complex on the surface of APCs. In addition, the parasite mitogens employed in this invention do not require activation of the APCs as manifested by the expression of: (1) adhesion molecules that promote the physical interaction between APCs and T cells; (2) membrane bound growth/differentiation molecules (co-stimulators) that promote T cell activation; or (3) soluble cytokines, such as IL-1 and TNF, as is required in the process for presenting antigens.

The mitogen employed in this invention is also distinguished from a "superantigen", which is a substance that can stimulate all of the T cells in an individual that express a particular set or family of V_{β} T cell receptor genes. Superantigens are typically bacterial and viral products, and can either be soluble or cell-bound. They do not require degradation to peptides. Superantigens are typically presented to the T cell receptor (TCR) on MHC molecules; however, they do not require processing by antigen presenting cells (APC), as do antigens, in order to be presented.

Thus, used herein, the term "mitogen" refers to a polyclonal activator that has the capacity to bind to and to trigger proliferation or differentiation of B lymphocytes, T lymphocytes, or mixtures thereof. Lymphocyte proliferation or transformation is the process whereby new DNA synthesis and cell division takes place in lymphocytes after a stimulus of some type, resulting in a series of changes. The lymphocytes increase in size, the cytoplasm becomes more extensive, the nucleoli are visible in the nucleus, and the lymphocytes resemble blast cells. The term blast

transformation is also sometimes applied to this process. Mitogens can induce proliferation in normal cells in culture. Activation of the lymphocytes thus can be characterized by transformation of the lymphocytes into blast cells, synthesis of DNA, cell division, increased production of immunoglobulins, or increased cytokine production. More particularly, the mitogens employed in this invention can stimulate whole classes of lymphocytes in this manner, and not just clones of particular specificity. The mitogens employed in this invention function, therefore, in a manner similar to the effects produced by lipopolysaccharide (LPS) on B cells, or lectins, concanavalin A (ConA), and phytohemagglutinin (PHA) on T cells.

With these phenomena in mind, the expression "parasite mitogen", as used herein, means at least one protein or polypeptide found in a parasite, wherein the protein or polypeptide is capable of provoking non-specific polyclonal activation of B lymphocytes, T lymphocytes, or mixtures thereof, in an *in vitro* culture of the lymphocytes in the manner similar to that just described. The protein or polypeptide comprising the parasite mitogen can be in glycosylated or non-glycosylated form. The parasite mitogen can be in natural or recombinant form.

The term "recombinant" as used herein means that a protein or polypeptide employed in the invention is derived from recombinant (e.g., microbial or mammalian) expression systems. "Microbial" refers to recombinant proteins or polypeptides made in bacterial or fungal (e.g., yeast) expression systems. As a product, "recombinant microbial" defines a protein or polypeptide produced in a microbial expression system, which is essentially free of native endogenous substances. Proteins or polypeptides expressed in most bacterial cultures, e.g. *E.*

coli, will be free of glycan. Proteins or polypeptides expressed in yeast may have a glycosylation pattern different from that expressed in mammalian cells.

The parasite mitogen employed in this invention can be in isolated or purified form. The terms "isolated" or "purified", as used in the context of this specification to define the purity of protein or polypeptide compositions, means that the protein or polypeptide composition is substantially free of other proteins of natural or endogenous origin and contains less than about 1% by mass of protein contaminants residual of production processes. Such compositions, however, can contain other proteins added as stabilizers, excipients, or co-therapeutics. The parasite is isolated if it is detectable as a single protein band in a polyacrylamide gel by silver staining.

Evaluation of lymphocyte proliferation can be quantitated in an assay of proliferative activity. For example, a radiolabelled precursor of DNA (usually tritiated thymidine) can be added to a culture medium and the amount of radioactivity incorporated into the cells subsequently detected. A suitable assay involves the *in vitro* culture of a lymphocyte population in the presence or absence of a mitogen for various periods of time. The changes induced in the stimulated groups are compared with changes in unstimulated cell populations. Radiolabelled amino acids are convenient as they provide a means of quantitating the changes in a simple, reproducible manner. Thus, as used herein, the expression "assay of proliferative activity" means the following assay:

Assay of proliferative activity

In vitro proliferation is accomplished using freshly recovered splenocytes from BALB/c mice seeded at a density of 5×10^4 cells/well and incubated for 24, 48 and 72 h with increasing concentrations of total parasite supernatants or recombinant TcPa45 protein or other mitogen (0.07-200 $\mu\text{g/ml}$) with 0.5 $\mu\text{g/ml}$ of the HPLC fractions, or with the conventionally used mitogens concanavalin A (10 $\mu\text{g/ml}$) and lipopolysaccharide (5 $\mu\text{g/ml}$) in 5% FCS in RPMI-1640 complete medium. T-cell depletion is accomplished by incubating freshly recovered spleen cells for 30 min. at 37°C in the presence of monoclonal antibodies against Thy 1.2 and rabbit complement (Cedarlane, Le Perray en Yvelines, France). Analysis of proliferative activity of total splenocytes (5×10^4 cells/well) in the presence of 50 $\mu\text{g/ml}$ enzymatically active rTcPA45 or other mitogen is also compared with the proliferation obtained using the same amounts of rTcPA45 protein or other mitogen lacking racemase activity (by heating for 10 min. at 30°C or by long term storage at 4°C). Inhibition of proliferation is obtained by adding to the splenocyte cultures 50 $\mu\text{g/ml}$ rTcPA45 or other mitogen pre-incubated for 10 min. at 37°C with 1mM inhibitor, either specific (pyrrole-2-carboxylic acid) or nonspecific (iodoacetamide or iodoacetate). Competitive assays of proliferative activity by 50 $\mu\text{g/ml}$ rTcPA45 or other mitogen are done by adding increasing concentrations of specific substrates i.e., proline racemase substrates (L- or D-proline) for the mitogen rTcPA45 ranging from 3 mM to 50 mM. Controls include the incubation of splenocytes (5×10^4 cells/well) with substrate alone, i.e., 50 mM L- or D-proline in RPMI medium alone for proline racemase. Cultures are collected after a 16-hour pulse or 1 $\mu\text{Ci/well}$ ^3H -thymidine uptake

was determined in a beta-plate liquid scintillation counter (LKB-Wallac, Orsay, France). All data points are obtained in triplicate and the corresponding standard deviation is calculated.

This assay of proliferative activity is used to determine whether a substance is a parasite mitogen. This assay is also used to determine a sub-mitogenic amount of the parasite mitogen. The results obtained in a typical assay of proliferative activity for two different mitogens, A and B, are depicted in Fig. 13, which is not based on data actually obtained in this invention, but which is merely included for illustrative purposes to show the effects produced by mitogens after lymphocyte activation.

Moreover, while the identification of a parasite or virus mitogen for use in the invention is accomplished by use of the assay of proliferative activity, it will be understood that the results obtained with this assay can be followed by other methods of measuring activation of lymphocytes, such as by measuring immunoglobulin production and/or carrying out immunoglobulin specificity assays. The Elispot assay described hereinafter can be used for this purpose.

As used herein, the term "sub-mitogenic amount" means an amount of the parasite mitogen, which is less than an amount of the parasite mitogen that produces an increase in lymphocyte proliferation in the assay of proliferative activity. Thus, the sub-mitogenic amount can be easily determined by carrying out the assay of proliferative activity at several low dosages of the parasite mitogen and noting the dosage at which proliferative activity first increases. The sub-mitogenic amount is an amount below the dosage at which proliferative activity first increases.

The sub-mitogenic amount must also be sufficient to induce protective immunity against the parasite in a host to which the sub-mitogenic amount of the parasite mitogen is administered. As used herein, the term "protective immunity" refers to an adaptive (specific) immune response characterized by specificity and memory in the host to which the sub-mitogenic amount of the parasite mitogen is administered. The adaptive immune response once stimulated by an invading parasite will remember and respond more rapidly to infection so that no disease will occur or any disease that occurs following infection will be less severe as compared to a similar infection without prior immunization according to the invention. Thus, the protective immunity imparted by the method of the invention imparts protection from disease, particularly infectious disease, as evidenced by the absence of clinical indications of disease, or as evidenced by absence of, or reduction in, determinants of pathogenicity, including the absence or reduction in persistence of the infectious parasite or virus *in vivo*, and/or the absence of pathogenesis and clinical disease, or diminished severity thereof, as compared to individuals not treated by the method of the invention.

The determination of a sub-mitogenic amount can readily be understood by reference to Fig. 13, which depicts two dose-response curves for two different hypothetical mitogens A and B. A sub-mitogenic amount of mitogen A would be an amount below about dose "Y" in Fig. 13, while a sub-mitogenic amount of mitogen B would be an amount below dosage "X" in Fig. 13.

The TcPA45 protein of the invention is referred to as a "parasite mitogen" in a functional sense in that it is capable of activating a non-specific polyclonal response in lymphocytes in the assay of proliferative activity. TcPA45 itself might not be a mitogen, but may act through racemization or by binding to host molecules, which would be the primary mitogens. Regardless, the non-specific lymphocyte activation by TcPA45 in amounts exceeding a sub-mitogenic amount would insure evasion of the parasite at the very beginning of infection.

The evasiveness and diversity of parasites has made definitive treatment difficult. Presented here are methods and agents for preventing the spread of parasitic and viral infections in a host, such as a human. Examples of microorganisms against which the methods and agents of the invention are effective are the following.

Table 2. Immune dysfunctions observed after mitogen-induced polyclonal activation following infectious processes		
Microorganism	Target lymphocytes	Acute or progressive dysfunctions
<i>Actinomyces viscosus</i>	B	Immunosuppression
<i>African Swine Fever virus</i>	B	Immunosuppression
<i>Ascaris</i>	B	IgE secretin, allergy, cerebral granuloma
<i>Borrelia burgdorferi</i>	B	Autoimmune arthritis
<i>Candida albicans</i>	B	Granuloma formation, immunosuppression
<i>Chlamidia trachomatis</i>	B	Lymphocytosis, autoimmunity
<i>Entamoeba histolytica</i>	T	Immunosuppression, disabling colitis, liver abscess
<i>Escherichia coli</i>	B	Toxic shock syndrome, meningitis, neurological and systemic symptoms
<i>Leishmania donovani, L major</i>	B	Immunosuppression, autoimmunity
<i>Listeria monocytogenes</i>	B and T	Meningitis, immune complex formation and immunosuppression
<i>Mycobacterium tuberculosis</i>	T	Immunosuppression and autoimmune arthritis
<i>Plasmodium chabaudi, P. yoellii</i>	T	Immunosuppression, autoimmunity

<i>P. falciparum</i>	B and T	Immunosuppression, autoimmunity
<i>Salmonella paratyphi</i> , <i>S. typhimurium</i>	B	Lethal septicemia, vascular myocardial injuries, immunosuppression, autoimmunity
<i>Schistosoma mansoni</i> , <i>S. hematobium</i>	B	Immunosuppression, megasyndromes, granuloma
<i>Staphylococcus aureus</i>	B	Toxic shock syndrome, mastitis, immunosuppression
<i>Streptococcus intermedius</i> , <i>S. mutans</i>	B	Toxic shock syndrome, immunosuppression
<i>S. pyogenes</i>	B and T	Immunosuppression, autoimmunity
<i>Toxocara canis</i>	B	Eosinophilia, lung damage, ocular granuloma, vasculitis
<i>Toxoplasma gondii</i>	B and T	Encephalitis, myocarditis, immunosuppression
<i>Trypanosoma brucei</i>	B	Immunosuppression, glomerulonephritis and brain lesions
<i>T. congolense</i>	B	Immunosuppression
<i>T. cruzi</i>	B and T	Hypergammaglobulinemia, immunosuppression, autoimmune myocarditis, megasyndromes

These and other substances employed as mitogens in this invention are of parasitic or viral origin, are of natural or recombinant form, and are similarly capable of producing a polyclonal response in unselected lymphocytes in the assay of proliferative activity. More particularly, while this invention has been described with reference to parasite mitogens, it is equally applicable to viral antigens. Indeed, by substituting "viral mitogen" for the expression "parasite mitogen" and "virus" for "parasite" in the foregoing description, the full scope of the subject invention will be understood. In addition, the parasite mitogens employed in this invention will exemplified with reference to the mitogen TcPA45 protein of *T. cruzi*. While the

parasite mitogen TcPA45 is also a racemase, it will be understood that the parasite mitogens and virus mitogens can be employed in this invention may or may not possess racemase activity.

Following are additional examples of microorganisms against which the vaccination strategy of the invention is applicable: Infection with Epstein-Barr, influenza, herpes, and sendai viruses; non- or poorly pathogenic viruses, such as and African swine fever virus; and murine leukaemia virus. Both T-cell-dependent and independent polyclonal B-cell activation have also been described for a variety of protozoan parasite infections, and these infections can also be prevented or abated by the vaccination strategy of this invention. These microorganisms include *Plasmodium berghei*, *P. yoellii*, *P. chabaudi*., *P. falciparum* and *P. vivax*.

In practicing the method of the invention, the parasite or viral mitogen is administered to a host using one of the modes of administration commonly employed for administering drugs to humans and other animals. Thus, for example, the parasite or viral mitogen can be administered to the host by the oral route or parenterally, such as by intravenous or intramuscular injection. Other modes of administration can also be employed, such as intrasplenic, intradermal, and mucosal routes. For purposes of injection, the mitogens described above can be prepared in the form of solutions, suspensions, or emulsions in vehicles conventionally employed for this purpose.

It will be understood that the parasite and viral mitogens can be used in combination with other parasite or viral mitogens or other prophylactic or therapeutic substances. For example, mixtures of different parasite mitogens or mixtures of different viral mitogens can be employed in the method of the invention. Similarly, mixtures of parasite and viral mitogens can be employed in the same composition. The parasite and viral mitogens can also be combined with other vaccinating agents for the corresponding disease, such microbial immunodominant, immunopathological and immunoprotective epitope-based vaccines or inactivated attenuated, or subunit vaccines. The parasite and viral mitogens can even be employed as adjuvants for other immunogenic or vaccinating agents.

The parasite or viral mitogen is employed in the method of the invention in an amount sufficient to provide an adequate concentration of the drug to prevent or at least inhibit infection of the host *in vivo* or to prevent or at least inhibit the spread of the parasite or virus *in vivo*. The amount of the mitogen thus depends upon absorption, distribution, and clearance by the host. Of course, the effectiveness of

the parasite or viral mitogen is dose related. The dosage of the parasite or viral mitogen should be sufficient to produce a minimal detectable effect, but the dosage should be less than the dose that activates a non-specific polyclonal lymphocyte response as measured by the assay of proliferative activity previously described.

The dosage of the parasite or viral mitogen administered to the host can be varied over wide limits. The parasite or viral mitogen can be administered in the minimum quantity, which is therapeutically effective, and the dosage can be increased as desired up the maximum dosage tolerated by the patient. The parasite or viral mitogen can be administered as a relatively high sub-mitogenic amount, followed by lower maintenance dose, or the parasite or viral mitogen can be administered in uniform dosages.

The dosage and the frequency of administration will vary with the parasite or viral mitogen employed in the method of the invention. In the case of the TcPA45 parasite mitogen, the sub-mitogenic amount administered to a human can vary from about 50ng per Kg of body weight to about 1µg per Kg of body weight, preferably about 100ng per Kg of body weight to about 500ng per Kg of body weight. Similar dosages can be employed for the other parasite and viral mitogens employed in this invention but optimum amounts can be determined with a minimum of experimentation using conventional dose-response analytical techniques or by scaling up from studies based on animal models of disease.

The term "about" as used herein in describing dosage ranges means an amount that is equivalent to the numerically stated amount as indicated by the induction of protective immunity in the host to which the parasite or viral mitogen is administered, with the absence or reduction in the host of determinants of pathogenicity, including an absence or reduction in persistence of the infectious parasite or virus *in vivo*, and/or the absence of pathogenesis and clinical disease, or diminished severity thereof, as compared to individuals not treated by the method of the invention.

The dose of the parasite or viral mitogen is specified in relation to an adult of average size. Thus, it will be understood that the dosage can be adjusted by 20-25% for patients with a lighter or heavier build. Similarly, the dosage for a child can be adjusted using well known dosage calculation formulas.

The parasite or viral mitogen can be used in therapy in the form of pills, tablets, lozenges, troches, capsules, suppositories, injectable in ingestable solutions, and the like in the treatment of cytopathic and pathological conditions in humans and susceptible non-human primates and other animals.

Appropriate pharmaceutically acceptable carriers, diluents, and adjuvants can be combined with the parasitic and viral mitogens described herein in order to prepare the pharmaceutical compositions for use in the treatment of pathological conditions in animals. The pharmaceutical compositions of this invention contain the active mitogens together with a solid or liquid pharmaceutically acceptable nontoxic carrier. Such pharmaceutical carriers can be sterile liquids, such as water and oils, including those of petroleum, animal, vegetable, or synthetic origin. Examples of suitable liquids are peanut oil, soybean oil, mineral oil, sesame oil and the like.

Water is a preferred carrier when the pharmaceutical composition is administered intravenously. Physiological solutions can also be employed as liquid carriers, particularly for injectable solutions.

The ability of the vaccines of the invention to induce protection in a host can be enhanced by emulsification with an adjuvant, incorporation in a liposome, coupling to a suitable carrier, or by combinations of these techniques. For example, the vaccines of the invention can be administered with a conventional adjuvant, such as aluminum phosphate and aluminum hydroxide gel. Similarly, the vaccines can be bound to lipid membranes or incorporated in lipid membranes to form liposomes. The use of nonpyrogenic lipids free of nucleic acids and other extraneous matter can be employed for this purpose.

Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatine, malt, rice, flour, chalk, silica gel, magnesium carbonate, magnesium stearate, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene glycol, water, ethanol, and the like. These compositions can take the form of solutions, suspensions, tablets, pills, capsules, powders, sustained-release formulations and the like. Suitable pharmaceutical carriers are described in "Remington's Pharmaceutical Sciences" by E. W. Martin. The pharmaceutical compositions contain an effective therapeutic amount of the parasite or viral mitogen together with a suitable amount of carrier so as to provide the form for proper administration to the host.

The host or patient can be an animal susceptible to infection by the parasite or virus, and is preferably a mammal. More preferably, the mammal is selected from

the group consisting of a human, a dog, a cat, a bovine, a pig, and a horse. In an especially preferred embodiment, the mammal is a human.

Another aspect of the invention includes administering nucleic acids encoding parasite and/or viral mitogens with or without carrier molecules to an individual.

Those of skill in the art are cognizant of the concept, application, and effectiveness of nucleic acid vaccines (e.g., DNA vaccines) and nucleic acid vaccine technology as well as protein and polypeptide based technologies. The nucleic acid based technology allows the administration of nucleic acids encoding parasite and/or viral mitogens, naked or encapsulated, directly to tissues and cells without the need for production of encoded proteins prior to administration. The technology is based on the ability of these nucleic acids to be taken up by cells of the recipient organism and expressed to produce a mitogen to which the recipient's immune system responds. Such nucleic acid vaccine technology includes, but is not limited to, delivery of naked DNA and RNA and delivery of expression vectors encoding the parasite or viral mitogen. Although the technology is termed "vaccine", it is equally applicable to immunogenic compositions that do not result in a complete protective response. Such partial-protection-inducing compositions and methods are encompassed within the present invention.

Although it is within the present invention to deliver nucleic acids encoding the parasite or viral mitogens as naked nucleic acids, the present invention also encompasses delivery of nucleic acids as part of larger or more complex compositions. Included among these delivery systems are viruses, virus-like particles, or bacteria containing the nucleic acids encoding the parasite or viral mitogen. Also, complexes of the invention's nucleic acids and carrier molecules with

cell permeabilizing compounds, such as liposomes, are included within the scope of the invention. Other compounds, such as molecular vectors (EP 696,191, Samain et al.) and delivery systems for nucleic acid vaccines are known to the skilled artisan and exemplified in, for example, WO 93 06223 and WO 90 11092, U.S. 5,580,859, and U.S. 5,589,466 (Vical patents), which are incorporated by reference herein, and can be made and used without undue or excessive experimentation.

Results indicate that intramuscular DNA vaccination protocols using pcDNA3 vector containing the TcPA45 gene, with or without the fragment encoding the signal peptide, are able to induce a decrease of 85% in parasitemia levels after challenge with infective forms of the parasite. Moreover, even higher levels of parasitemia control resulted when sub-mitogenic doses of the active rTcPA45 protein were injected intraperitoneally 2 weeks before infective challenge. These observations support the use of this molecule as a drug and/or immunomodulator target.

This invention further contemplates:

1. Any molecular modification of the gene or a fragment of the gene encoding for a racemase/mitogen that leads to the inhibition of the expression of the protein by the parasite or virus (gene knock out), and further utilization of parasites or viruses lacking those activities *in vivo* aiming at immunoprotective responses.
2. Any molecular modification of the gene or a fragment of the gene encoding for a racemase/mitogen that leads to the hyperexpression of the protein by the parasite or virus (gene transgenesis), and further utilization of the parasite or virus to produce high amounts of the protein aiming at producing high amounts of the native protein.
3. Any molecular modification of the gene or a fragment of the gene encoding for a racemase/mitogen that leads to an attenuation of parasite or virus infectivity, or interaction with a host cell, and further injection of the parasites or viruses *in vivo* aiming at immunoprotective responses.
4. Any molecular modification (for instance directed mutagenesis) of the protein or of its active site that leads to the inhibition of its enzymatic or its mitogenic activity and further injection of mutated parasites or viruses *in vivo* aiming at immunoprotective responses.
5. Use of any molecular or biochemical modification of the enzymatic activity of the racemase (inhibition of the active site) aiming at developing specific immunotherapy.
6. Any molecule or compound that inhibits the enzymatic activity of the protein aiming at developing a drug against parasite or virus infection or specific treatment of parasitic or viral disease.

An example of the application of this technology to the invention follows:

The catalytic site of TcPA45 protein responsible for racemase activity is identified by the boxed region in Fig. 2. The catalytic site comprises the amino acids SPCGT. Inhibition of racemase activity, and the consequent loss in infectivity, can be accomplished by altering this catalytic site in the protein or altering the corresponding nucleotides in the gene encoding the protein. A target for alteration would be the cysteine residue. For example, changing the cysteine residue to serine does not significantly alter the charge on the molecule, but diminishes racemase activity. The catalytic site can be altered in other ways, such as by the addition, deletion, or substitution of another moiety for one of the moieties in the protein or the nucleic acid encoding the protein so that secondary and tertiary structures are not materially altered, binding sub-units are not affected, but racemase activity is diminished or totally lost.

Plasmids containing the polynucleotides from *T. Cruzi* have been deposited at the Collection Nationale de Cultures de Microorganismes ("C.N.C.M.") Institut Pasteur, 28, rue du Docteur Roux, 75724 Paris Cedex 15, France, as follows:

<u>Plasmid</u>	<u>Accession No.</u>	<u>Deposit Date</u>
DH5 α -pTc45MIT (239 bp)	I-2221	June 9, 1999
DH5 α -pTc45MIT (1335 bp)	I-2344	October 29, 1999.

This invention will now be described with reference to the following Examples.

EXAMPLE 1 - Mice and Parasites.

Trypanosoma cruzi clone CL Brener was used throughout this work. Epimastigotes were maintained by weekly passage in liver infusion tryptose medium.

In vitro metacyclogenesis was performed in a protein free defined medium at 27°C, as previously described¹¹. Male euthymic or athymic BALB/c mice 8 weeks of age were purchased from Charles River Laboratories (Saint Aubin les Elbeuf, France). Male C3H/He mice 8 weeks of age from our animal facilities were also used.

EXAMPLE 2 - Protein fractionation.

40 liters of culture supernatants from metacyclic forms, maintained for an additional 96h at 37°C, were concentrated by vacuum dialysis and dialyzed against buffer A. HPLC was performed using a weak anion exchanger column POROS HQ-10 (Perspective Biosystems) at a flow rate of 1ml/min according to the following program: a) 10 min with buffer A; b) 30 min linear gradient from buffer A to B; c) 5 min linear gradient from buffer B to C; and d) 5 min with buffer C. One ml fractions were collected, frozen at -80°C, lyophilized and reconstituted in H₂O or in non-supplemented RPMI medium for *in vitro* proliferation assays. (Buffers used: A: 5 mM NH₄-acetate, pH 8. B: 1M NH₄-acetate, pH 8. C: 1M NaCl/1 M NH₄-acetate, pH 8). Fractions 1 ml in volume were collected, frozen at -80°C, lyophilized and reconstituted in water or in non-supplemented RPMI medium for *in vitro* proliferation assays. SDS-PAGE analysis used standard techniques.

EXAMPLE 3 - Generation of peptides and amino acid sequence analysis.

HPLC fractions 22 and 23 were pooled and fractionated by 8% SDS-PAGE. After amino black staining, the 45 kDa protein band was cut out, in-gel digested with trypsin, and submitted to reverse phase HPLC to separate peptides. Automated Edman degradation sequence analysis was performed in the Laboratoire de Microséquençage de Protéines of the Pasteur Institut.

EXAMPLE 4 - RNA preparation, reverse transcription, PCR, and cloning.

RNA was extracted from trypomastigote forms obtained from Vero cells, using TRIzol LS reagent (Gibco) following manufacturer's instructions. Two µg from this RNA were reverse transcribed in 20 µl with Superscript II (Gibco) using anti-sense degenerate primer 5'TTICCRAADATIACIACGTT3' [SEQ ID NO: 12] designed from peptide 5.

PCR reaction was performed on 5 µl of cDNA using Taq polymerase (Perkin Elmer) or Pfu DNA polymerase (Stratagene). The following PCR conditions and primers were used: 1. *TcPA45* gene fragment (239 bp) 30s at 94°C, 45s at 45°C, 30s at 72°C for 30 cycles followed by 10 min at 72°C; degenerate primers: corresponding to peptide 4 (forward) 5'ATHGCITTYGGIGGIAAYTTT3' [SEQ ID NO: 13] and to peptide 5 (reverse): 5'TTICCRAADATIACIACGTT3' [SEQ ID NO: 14]. (D for A, G or T; H stands for A, C or T; M for A or C; I for inosine; R for A or G; Y for C or T). 2.

The *TcPA45* coding sequence (from codons 30 to 423) was amplified using 45s at 94°C, 45s at 50°C, 3 min at 72°C for 20 cycles, with primers 5'CTCTCCCATGGGGCAGGAAAAGCTTCTG3' [SEQ ID NO: 15] and

5'CTGAGCTCGACCAGATCTATCTGC3' [SEQ ID NO: 16]. PCR products were purified with Qiagen PCR extraction kit and cloned into pCR II-TOPO vector using the TOPO-TA cloning kit (Invitrogen) following manufacturer's instructions.

EXAMPLE 5 - Automated sequencing.

Lambda phage and plasmid DNA were prepared using standard techniques, and direct sequencing was performed using Big Dye Terminator kit (Perkin Elmer) following manufacturer's instructions. Extension products were run for 7 h in an ABI 373B automated sequencer. Primers internal to the sequence have also been used for sequencing.

EXAMPLE 6 - Genomic library screening.

A genomic library of *T. cruzi* CL-Brener constructed in phage lambda Fix II (from Dr. E. Rondinelli, UFRJ, Brazil) was screened using a ³²P labelled 239 bp PCR product as a probe. Hybridization was performed using standard conditions. Filters were scanned using a PhosphorImager scanning unit (Molecular Dynamics). Positive phages were identified and phage DNA was prepared using standard procedures.

EXAMPLE 7 - Expression constructs and recombinant protein expression.

The PCR product encoding the TcPA45 gene fragment starting at codon 30 was cloned in frame with a C-terminal 6x histidine tag into the pET28b(+) expression vector (Novagen). The soluble recombinant protein was produced in *E. coli*, and the soluble fraction was purified using a Ni²⁺ column (Novagen) following manufacturer's instructions.

EXAMPLE 8 - Racemase activity.

Demonstration of the racemase enzymatic activity of rTcPA 45 used a polarimeter. Buffer Na-acetate (pH 6), reaction Vol. 500 μ l. Optimum pH was found to be 6 and the temperature 37°C.

3 μ g of rTcPA45 was diluted in 500 μ l of buffer (0.2M NaOAc, 20mM β -mercaptoethanol, pH6.0) containing different concentrations of the substrate (10-80 mM or L- or D-proline). (See Figure 4c.) The reaction was incubated for 30-60 minutes at 37°C in a water bath, followed by heat inactivation of the enzyme for 10 minutes at 80°C. Each sample is diluted to 1.5 ml with water and the mixture submitted to measurement of optical rotation using a polarimeter, at 365nm. A sample without enzyme is used as a control. For optimal pH determination, buffer systems with NaOAc, phosphate, and Tris were used with pH ranging from 4.0 - 8.2. Combined results are shown in Figure 4e. Curve of temperature was performed between 27°C to 47°C.

EXAMPLE 9 - Mitogenic activity.

Analysis of mitogenic proliferative activity of spleen cells *in vitro* with rTcPA45 (His/tag) protein.

The figure [Figure 4] is representative of 3 experiments using different mouse strains (C57BL/6, BLAB/c and C3H:HeJ, the latter a LPS-nonresponsive strain). Similar results were obtained with the 3 strains, including C3H/HeJ, showing that our preparation is not contaminated by bacterial LPS. The proliferation is dose-dependent and presents a bi-modal pattern as already observed with total culture supernatants used to purify the Tc45 protein. Best cell concentration = 5×10^4 cells/well.

5×10^5 naive spleen cells/well (96 well plate) were stimulated *in vitro* with different doses of rTcPA45 (ranging in Figure 4b from 0.8 to 200 $\mu\text{g/ml}$ final) for 24, 48 and 72h, at 37°C , 5% CO_2 . Cultures were pulsed with ^3H -Thymidine (1 $\mu\text{Ci/well}$) for 16-18h before harvesting. ^3H -thymidine incorporation was obtained after counting using a beta-plate. Results present arithmetic means of c.p.m. (counts/minute) from 6 wells/dose of rTcPA45 or wells containing medium alone (+/- SD of the means). (Figure 4b)

EXAMPLE 10 - Mitogenicity of rTcPA45 *in vivo* assessed by ELISPOT.

BALB/c mice were injected or not with 50 μg of rTcPA45 (i.p.), and spleen cells assayed day 7 after injection. Results represent total numbers of spleen cells, total number of B cells producing IgM, IgG2a, or IgG2b isotypes, and total numbers of isotype-producing B cells specific of the protein.

TABLE 1

ELISPOT assay 7 days after i.p. injection of rTc45MIT

Total Number of Ig-producing B cells

	IgM-producing cells	IgG2a-producing cells	IgG2b-producing cells
Non-injected	163000 \pm 25456	3650 \pm 636	4300 \pm 142
PTc45MIT (50 μ g/mouse)	371666 \pm 94495	9866 \pm 3000	14633 \pm 3287

Total Number of Ig-producing B cells ANTI-rTc45MIT

	IgM-B cells anti 45	IgG2a-B cells anti 45	IgG2b-B cells anti 45
Non-injected	none	none	none
PTc45MIT (50 μ g/mouse)	84	none	none

It is observed 1) a 2 fold increase in total spleen cell numbers after 7 days of rTcPA45 injection, 2) 3-6 fold increase in total numbers of Ig-secreting cells, and 3) less than 0.5% of IgM secreting cells are directed to the injected protein, characterizing a mitogenic stimulation of B cells.

Mitogenicity of the rTcPA45 in vivo (assay)

Mice were injected or not with 50 μ g of iTcPA45 (i.p.). 7 days later, spleens were removed and cell suspensions were prepared and counted. Numbers of Ig-secreting cells (total or specific to rTcPA45) were determined by Elispot assay, as follows:

ELISPOT ASSAY

Flat bottomed 96 well ELISA plates, were coated with either goat anti-mouse Ig or with rTcPA45 and incubated at 4°C overnight. Plates were blocked with PBS-gelatin, washed with PBS-Tween and with RPMI medium. 100 μ l of

different spleen cell concentrations per well (in RPMI containing 2% FCS) were incubated for 8 hours at 37°C, 5% CO₂. A row in the plate containing serial dilutions of purified immunoglobulin (IgM, IgG2a, or IgG2b) was used as standard. After lysis of the cells with H₂O-Tween, plates were then washed with PBS-Tween and incubated with respective biotin-labeled antibodies directed to (IgM, IgG2b, or IgG2a isotypes), overnight at 4°C. After washing, plates were incubated for 45-60 minutes with avidin-alkaline phosphatase and further incubated with substrate (2-amino-2-methyl-propanol buffer containing BCIP) 2-4 hours, 37°C (until the spots are "dark" blue). Spots correspond to immunoglobulin-producing B-cell (of a particular isotype) directed to the coated antigen (here: goat anti-mouse immunoglobulins or rTcPA45). Spots are then counted and numbers corrected to total number of spleen cells according to the dilution. (See Table I and Figure 7.)

EXAMPLE 11 - DNA vaccination

8 week old BALB/c mice (5 mice/group) were injected (i.m.) once, or 3 times (interval of 3 weeks) with the different constructions (100µl plasmid/femoral quadriceps), as follows:

- a) controls : saline and rTc24
- b) Vectors (pcDNA3 and VR1020, which is described by R. Amasamy et al., *Biochemica Biophysica Acta* 1998, Vol. 1453, pp. 1-13) of DNA vaccination containing different constructs : Long, containing the complete sequence of rTcPA45 gene; Short, containing the sequence of rTcPA gene without the signal peptide,

*VR1020 vector contains an additional signal peptide (tissue Plasminogen Activating factor, TPA).

c) empty vectors

Mice were challenged 4 weeks after the last injection with 10^4 infective forms of the parasite/mouse, and the parasitemia was scored during 35 days. Serum samples were collected before challenge and assayed by Western blot against the recombinant protein.

It is worth noting that BALB/c mice were treated by almost 2 months (9 weeks) to follow the vaccination protocol and were challenged at 21 week old. It is well known that over 10 week old mice are resistant to the experimental infection with *Trypanosoma cruzi* and no mortality is observed.

The results using this vaccination protocol revealed that 3 injections of pcDNA3 containing either the Short or Long constructs, or just 1 injection of the same vector with the Short construct is able to reduce by more than 50% the parasitemia levels. Titres of total immunoglobulins anti-rTcPA45, 4 weeks after the last plasmid injection : controls (saline and empty vectors) : 1 :100; both constructs in pcDNA3 (Short and Long) : 1 :2000, and respectively 1/1000 and 1 :2000 for VR1020 containing the Long and Short sequences. (See Figure 8.)

EXAMPLE 12 - Southern, Western and Northern blots.

Mice were immunized intrasplenically with 10 ng protein and were boosted every 3 weeks with 1 µg of the same preparation for 2 months to obtain polyclonal serum containing rTcPA45-specific antibodies. Total, soluble and insoluble sonic extracts, or culture supernatants from the different parasite forms were purified and

separated by 8-10% SDS-PAGE, and proteins were electrophoretically transferred to nitrocellulose membranes. Membranes were saturated with Tris-buffered saline and milk, incubated with polyclonal serum against rTcPA45 and developed with peroxidase-labeled secondary antibody using an ECL kit (Amersham, Orsay, France). *T. cruzi* genomic DNA (10 µg) was digested with restriction enzymes (*Bam*HI, *Bgl*II, *Sal*I, *Taq*I and *Pst*I), separated by 0.8% agarose gel electrophoresis and transferred to Hybond N+ followed by hybridization of the membrane with a ³³P-dATP-labeled probe covering the TcPA45-coding sequence. Total RNA was prepared from epimastigote, metacyclic and trypomastigote forms of the parasite by conventional methods. For northern blot analysis, 20 µg epimastigote RNA was transferred to Hybond N+ membranes, then hybridized with single-stranded DNA complementary to the *TcPA45* gene transcript, labeled with α-³²P-dCTP.

Transcript analysis through reverse transcription and PCR. Total RNA (1 µg) from epimastigote, metacyclic and trypomastigote forms of the parasite were used to synthesize specific first-strand cDNA by using oligonucleotide R300-45 (5'-TCCGTATCCATGTCGATGC-3') [SEQ ID NO:24], located about 240 nucleotides downstream from the first ATG start codon, followed by PCR amplification using R300-45 and an oligonucleotide corresponding to part of the *T. cruzi* spliced leader sequence (5'-TATTATTGATACAGTTTCTG-3') [SEQ ID NO:25]. An internal TcPA45 fragment of about 170 bp was then amplified using R300-45 and the oligonucleotide HI-45 (5'-CTCTCCCATGGGGCAGGAAAAGCTTCTG-3') [SEQ ID NO:26] to demonstrate the presence of Tc45 transcript in each of the life stages analyzed.

EXAMPLE 13 - Immunofluorescence.

Cellular localization of TcPA45 protein in epimastigote, metacyclic and bloodstream forms of the parasite was demonstrated by indirect immunofluorescence using polyclonal mouse serum against rTcPA45 (described above) followed by 4 µg/ml Alexa 488™ goat antibody against mouse IgG (H+I), F(ab')₂ fragment conjugate (Molecular Probes-Interchim, Montlucon, France), compared with control staining using Alexa 488™ F(ab')₂ fragment conjugate alone or after incubation of the parasites with chronic serum obtained from mice infected for 8 months.

EXAMPLE 14 - Racemization assays.

The percent of racemization of different concentrations of L-proline, D-proline, L-hydroxy-proline and D-hydroxy-proline substrates was calculated by incubating a 500-µl mixture of 3 µg TcPA45 and 10-80 mM substrate in 0.2 M sodium acetate and 25 mM β-mercaptoethanol, pH 6, for 30 min at 37°C. The reaction was stopped by incubation for 10 min at 80°C. Water (1 ml) was then added, and the optical rotation was measured in polarimeter 241 MC (Perkin Elmer, Montignyle Bretonneux, France) at a wavelength of 365 nm, in a cell with a path length of 10 cm. The percent inhibition of racemization of 80 mM L-proline was determined in the presence of different concentrations of several specific and nonspecific inhibitors ranging in concentration from 6 mM to 100 mM. The percent racemization of 80 mM L-proline as a function of pH was determined using 0.2 M sodium acetate, potassium phosphate and Tris-HCl buffers containing 25 mM β-mercaptoethanol;

reactions were incubated for 30 min at 37°C as described above. All reagents and inhibitors were purchased from Sigma.

Accession numbers. The GenBank accession number of *T. cruzi* TcPA45 is AF195522. The EMBL accession number of *C. sticklandii* is E10199.

EXAMPLE 15

rTcPA45 is a B cell mitogen from a pathogenic trypanosome and a novel eukaryotic proline racemase. Both mitogenic and racemase activities seem to be linked and dependent on the integrity, or the availability of the enzyme active site. Specific inhibition of the active site (using specific or non-specific proline racemase inhibitors), or the active site occupancy by the substrate (competition assays using L- or D- proline), respectively, abolish or decrease B-cell mitogenic properties of rTcPA45.

As with classical B cell mitogens, rTcPA45 mitogenicity is dose-dependent (Fig. 1). As for classical B cell mitogens, mouse specific immune responses directed to mitogenic rTcPA45 is indeed possible (Western blot Figure 12) following a protocol of immunization which consisted of with 1 injection (i.p.) of a sub-mitogenic (10 ng/mouse) dose of the protein followed by an additional boost with a mitogenic dose of rTcPA45 (50 µg/mouse) one week later.

Protocol

5 groups of 6 week old Balb/c mice (3 mice per group) were immunized as follows :

T = non immunized.

Vide = immunized with 50 µg of empty PcDNA3 vector, i.m. per 3 week (twice).

Court = immunized with 50 µg of PcDNA3 - short (TcPA45 short), i.m. per week (twice).

I. S. = immunized with 10 ng of rTcPA45 i.s. (first week) and 50 µg rTcPA45 i.p. (second week).

I.P. = immunized with 10 ng of rTcPA45 i.p. (first week) and 50 µg rTcPA45 i.p. (second week).

Sera from individual mice were collected and analyzed by Western blot:

80 µg of rTcPA45 were loaded onto 0.8 % SDS-page gels and transferred to Hybond membranes.

Sera were individually tested at 1/400 dilution and reactivity against rTcPA45 revealed with anti-IgG mice immunoglobulins using chemoluminescence.

Figure 12 (J-14) represents individual serum reactivity before immunization before challenge (J0), and after 10 days (J10) or 21 days (J21) of challenge.

Challenge consisted of i.p 10000 parasites.

Additionally, rTcPA45 molecule can be considered as a target for vaccination strategies, since the previous Example using DNA-vaccination protocols (intramuscular injections of DNA-vectors containing the *Tc45 gene*) was able to reduce by 85% the parasitemia levels after an infectious challenge with the parasite. (See Fig. 14a.)

Moreover, experiments showed that higher levels of parasitemia control (90-95%) was obtained when the submitogenic protocol of immunization described above was followed by an infectious challenge (10^4 parasites/mouse). These additional results do support the claim that mitogenic moieties are potential targets for vaccination approaches against *Trypanosoma cruzi* infection. (See Fig. 14(b).)

* * *

In summary, we have investigated the mechanisms and consequences of polyclonal lymphocyte activation using the experimental model of Chagas disease, caused by the protozoan parasite *T. cruzi*. As in other infectious processes, this disease involves extensive B- and T-cell activation, hypergammaglobulinemia, and the establishment of chronic autoimmunity affecting the heart and the digestive tract. *Trypanosoma cruzi* infection induces a lymphocyte blast transformation of a magnitude that is similar to or higher than that induced by the classic polyclonal activator LPS.

This invention involves the use of a parasite or viral mitogen as a vaccinating agent without lymphocyte polyclonal activation that inhibits a protective host specific immune response to the parasite or virus. This invention not only prevents infection by the parasite or virus, but also avoids the negative consequences of such infection (immunosuppression, persistent infection, and susceptibility to immunopathology and autoimmune phenomenon). This invention thus addresses the correlation of polyclonal lymphocyte activation of the immune system after infections with poor specific responses and severe immunosuppression to autologous or unrelated challenges through an effective vaccination strategy.

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